



# Biodiversity of Mushrooms in the Long Island Pine Barrens Region



James Peperno and Jalal Sawas  
Shoreham-Wading River High School



## Abstract

DNA was extracted from mushrooms collected from the campus of the Shoreham-Wading River High School, identified by physical characteristics and then analyzed to confirm genetic identity. Species identified include *Laccaria trullisata*, *Tricholoma sp.* and *Neofavolus alveolaris*.

## Introduction

Fungi are a common species found in suburban areas, especially fruiting bodies such as mushrooms. While typically considered a nuisance, often removed and discarded, some of these fungi are essential to maintaining a healthy environment. Certain fungi can also help with human health in some ways, as well as being an indicator of a healthy forest environment. This study focuses on the Shoreham Wading-River community pond, where there are an abundance of mushrooms. Originally designed as a runoff station, the pond has the potential to have heightened amounts of trace metals such as: Fe, Mn, Zn, Cu, Pb, and Cr, due to the close proximity to a heavily trafficked road. Car exhaust is normally made up of primarily chromium, zinc, and iron and these chemicals, emitted as particulates, can settle out into the water and soil surrounding the pond (Pereira 2007). The presence of mushrooms such as the *Russula Emetica* or other genetically similar species would potentially reduce the chance of mutated organisms in and around the pond. To help facilitate the creation of a healthy pond ecosystem, identification of the species of mushrooms surrounding the pond will be genetically identified, allowing for the selection of species that are more efficient in their removal of metals from the soil components.



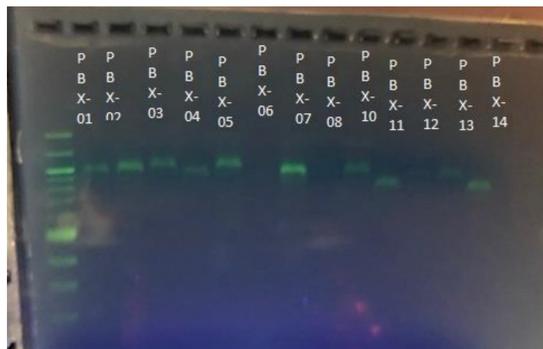
Collecting mushroom samples



## Materials and Methods

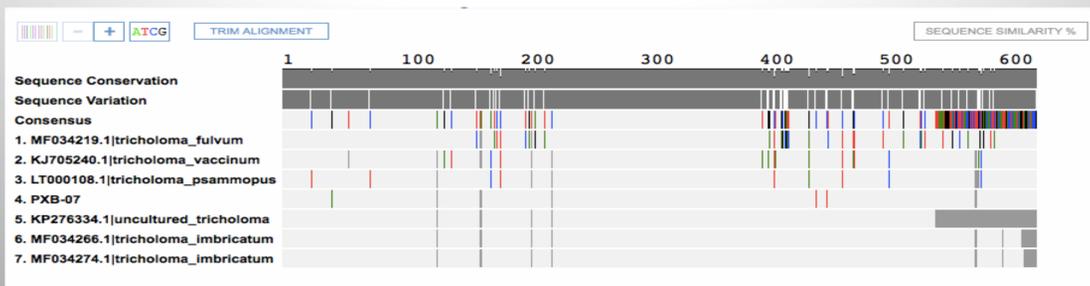
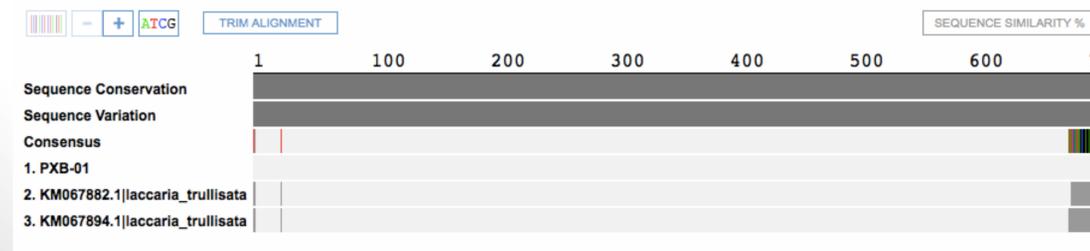
In this study, samples were collected in the Pine Barren region of Long Island, using a knife to remove mushrooms from their original location. Samples were photographed where they were found and growth habit was noted. Once removed from the site, samples were stored at 4°C. Sample species were taxonomically identified using Field Guide to Common Macrofungi in Eastern Forests and Their Ecological Functions.

As per the Barcode Long Island DNA extraction protocol, lysis solution was used in order to break down cell walls, then silica resin was added to bind DNA. After washing with wash buffer to remove impurities, DNA was released from the silica beads by eluting in distilled water. 50ul of this DNA solution was transferred to a new tube and stored at -20C until ready for PCR. In the PCR reaction we used the ITS1F primer, and PCR products were checked for amplified DNA using gel electrophoresis, and the resulting gel is shown below. Samples 1-5, 7, 10 and 11 were sent for sequencing.



Gel electrophoresis to confirm presence of DNA after PCR

## Results



## Discussion

None of the samples we identified taxonomically matched with the names provided by the BLAST search, but of the species that were sequenced, several did match our samples in appearance and therefore are believable specimen identification.

Mushrooms proved difficult to work with for extraction purposes, as their tissue is very dense and rubbery, but as evidenced from our electrophoresis gel post PCR, the majority of our samples amplified well despite this.

The diversity of mushrooms in the collection site in the Pine Barren region was fairly significant, as we were able to collect fourteen samples with very different looking appearances. Our next steps will be to analyze the soil around these collection locations as well as the tissues of the mushrooms to determine the effects of the mushrooms on the removal of metals from the soil environment.



Sample 1 *Laccaria trullisata*



Sample 11 *Neofavolus vaccinum*

## References

Busuioc, Gabriela, et al. "The Bioaccumulation and Translocation of Fe, Zn, and Cu in Species of Mushrooms from Russula Genus." *Environmental Science and Pollution Research International*, vol. 18, no. 6, 2011, pp. 890-6.  
Busuioc, G., Elekes, C.C., Stihl, C. et al. *Environ Sci Pollut Res* (2011) 18: 890. <https://doi.org/10.1007/s11356-011-0446-z>  
"Collecting Mushroom Samples for DNA Testing." *Penn State University*, news.psu.edu/gallery/486759/2017/10/06/collecting-mushroom-samples-dna-testing.  
Dighton, John "The New Jersey Pine Barrens." *Go to Pinelands Field Station*, pinelands.camden.rutgers.edu/about/the-pine-barrens/.  
*DNA Learning Center Barcoding 101*, www.dnabarcoding101.org/programs/bli/.  
ELEKES, Carmen, Gabriela BUSUIOC, & Gheorghe IONITA. "The Bioaccumulation of Some Heavy Metals in the Fruiting Body of Wild Growing Mushrooms." *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* [Online], 38.2 (2010): 147-151. Web. 4 Feb. 2019  
Michael E. Ostry Neil A. Anderson Joseph G. O'Brien "Field Guide to common Macrofungi in Eastern Forests and their Ecosystem Functions" 23 April 2010 United States Forestry Department PA  
"Submitting a Mushroom for Identification." *Submitting a Mushroom for Identification Horticulture and Home Pest News*, hortnews.extension.iastate.edu/pidc/mushroom  
Wu, Brian. "Fungi: Hazards and health applications." *Medical News Today*. MediLexicon, Intl., 14 Mar. 2017. Web. 26 Oct. 2018.  
<<https://www.medicalnewstoday.com/articles/158134.php>>

## Acknowledgements

We would like to thank Dr. Sharon Peperno, Jeffrey Petracca and Megan Capobianco from Barcode Long Island for all of their assistance and patience on this project.

Identification of sample mushrooms through taxonomic methods

Sample #	Common Name	Scientific name
1	Torn fibrecap	<i>Inocybe lacera</i>
2	Waxcap	<i>Hygrocybe sp.</i>
3	Yellow milkcap	<i>Lactarius chrysorrheus</i>
4	Sarahusokas	<i>Entoloma serrulatum</i>
5	America abrupt bulbod <i>Lepidella</i>	<i>Amanita polypyraxis</i>
7	Sarahusokas	<i>Entoloma serrulatum</i>
10	Panther cap	<i>Amanita pantherina</i>
11		<i>Gymnopus dichrous</i>